

Original Research Article

## ANTIMICROBIAL PROFILES OF UROPATHOGENS IN A TERTIARY CARE HOSPITAL OF NORTHWESTERN INDIA

Rupinder Bakshi<sup>1</sup>, Karashdeep Kaur<sup>2</sup>, Palika Sharma<sup>2</sup>, Jaspreet Kaur Boparai<sup>2</sup>,  
Satinder Kaur<sup>2</sup>, Ruchika<sup>1</sup>, Arunita<sup>1</sup>

<sup>1</sup>Department of Microbiology, GMC, Patiala

<sup>2</sup>Research Scientist, Viral Research & Diagnostic Laboratory, GMC Patiala

---

**Corresponding Author : Dr. Ruchika**

Junior Resident, Department of Microbiology, GMC, Patiala

E-mail: ruchika19t@gmail.com

---

### Abstract

**Background:** Urinary tract infections (UTIs) are extensively studied due to their high prevalence and potential severity, particularly concerning antimicrobial properties. This study aimed to evaluate the prevalence of common uropathogens and their antimicrobial resistance.

**Method:** A prospective cross-sectional study was conducted on a total of 16,077 urine samples at a tertiary care hospital in Patiala, Punjab, from January 2022 to December 2022 using standardized microbiological methods. The isolated microorganisms underwent antibiotic resistance testing using the Kirby-Bauer disk diffusion method.

**Results:** A total of 16,077 urine samples were analyzed, among gram negative microorganism *Esch.coli* (n=1962) was the most prevalent microorganism, followed by *Klebsiellapneumoniae* (n=722), *Pseudomonas aeruginosa* (n=572), *Acinetobacter Baumannii Complex*(451), *Citrobacter species* (n=132), *Proteus spp.* (n=96), and *Enterobacter spp.* (n=12). Among gram positive *Enterococcus spp.* (n=786) was most prevalent followed by *Staphylococcus aureus* (n=420), MRSA (n=30), Antibiotic profile data revealed that gram negative have high resistance rates towards Ampicillin, Ciprofloxacin and Amoxicillin-clavulanate and among gram positive resistance rate was higher in Ampicillin, Ciprofloxacin Amoxicillin-clavulanate, Gentamicin, Levofloxacin, Erythromycin and Netilmicin.

**Conclusion:** *Esch. coli* and *Enterococcus spp.* emerged as the predominant uropathogen, exhibiting a high level of antibiotic resistance against various antibiotics.

**Keywords:** Antibiotic resistance; Urinary tract infections; Uropathogens; gram negative; gram positive; Healthcare-associated infections

---

### Introduction:

Urinary tract infection (UTI) stands as the most prevalent bacterial infection, contributing to 25.0% of all reported infections [1]. It constitutes a significant cause of morbidity and ranks as the second most frequent reason for hospital visits. Approximately 35% of healthy women experience UTI symptoms at some point in their lives [2]. *Esch.coli*, a part of the normal bowel flora, is the primary causative agent, accounting for over 75% of UTI cases [3]. *Esch.coli* is increasingly implicated in both community-acquired and hospital-acquired

infections, facilitated by its ability to bind selectively to uroepithelial cells using P fimbriae [4]. UTIs present with symptoms such as fatigue, dysuria, urgency of urine, and urinary tract irritation. Prolonged hospital stay and indiscriminate antibiotic usage are the risk factors [5]. Other pathogens such as Gram-negative Enterobacteriaceae, Gram-positive *Enterococcus faecalis*, and *Staphylococcus saprophyticus* contribute to the remaining UTI cases. Although UTI affects both genders, it is more prevalent in women due to their shorter urethra and its proximity to the anus, with exceptions observed in

older men above the age of 60 with prostatic hypertrophy [6].

UTIs associated with *Esch. coli* lead to significant morbidity and long-term consequences, affecting approximately 10% of individuals globally [7]. These infections can manifest as asymptomatic or symptomatic, posing a substantial health burden. However, the effectiveness of antibiotics in controlling these infections is diminishing due to the widespread resistance of bacterial uropathogens to various antibiotics. Early detection of antibiotic resistance and the formulation of appropriate antibiotic policies are crucial for effective management in tertiary care settings [8].

Given the geographic variability in the prevalence of resistance strains, understanding the current levels of antimicrobial resistance among common urinary pathogens is essential [9]. Establishing institutional antibiotic policies can assist clinicians in selecting the most appropriate treatment strategies [10]. Therefore, this study aimed to investigate the prevalence and antibiotic sensitivity patterns of uropathogens in urine samples, with the findings intended to inform antibiotic policies and control measures.

#### Material and Methods:

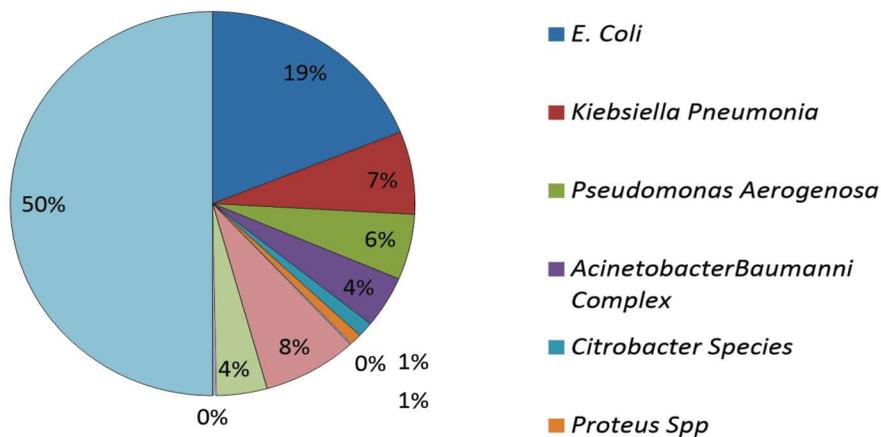
A retrospective cross-sectional study was conducted in the Department of Microbiology in a tertiary care hospital in Patiala from January 2022 to December 2022. A total of 16,077 urine samples were processed, including 577 samples from the ICU, 7500 samples from the outpatient unit, and 8000 samples from the inpatient unit. The conventional loop method, a semi-quantitative technique, was used to cultivate urine. The organisms isolated from urine culture were identified by standard methods<sup>1</sup>. The antibiotic sensitivity test was done on Mueller-Hinton agar by Kirby-Bauer disc diffusion test as per Clinical and Laboratory Standard Institute (CLSI) guidelines<sup>8</sup>. Antibiotics used for gram negative organism were Ampicillin (10µg), Amikacin (30µg), Gentamicin (10µg), Ciprofloxacin (5µg), Levofloxacin (5µg), Ofloxacin (5µg), Norfloxacin (10µg), Ceftazidime (30µg), Cefotaxime (30µg), Ceftriaxone

(30µg), Cefepime (30µg), Piperacillin- Tazobactam (100/10µg), Nitrofurantoin (300µg), Cotrimoxazole (25µg), Imipenem (10µg), Meropenem (10µg). For gram positive microorganism Ampicillin (10µg), Amoxycillin (30 µg), Amoxy-Clav (20/10µg), Erythromycin (15µg), Clindamycin (2µg), Netilmicin (30µg), Linezolid (30µg) and Vancomycin (30µg) antibiotics were used. If an isolate was discovered to be resistant to three or more antibiotics from various classes or groups of antibiotics, it was deemed multidrug resistant (MDR). Bacterial isolates were identified using standard microbiological protocols, and antibiotic susceptibility testing was performed using the Kirby-Bauer disk diffusion method. Various antibiotics were tested, and zone sizes were interpreted based on established criteria.

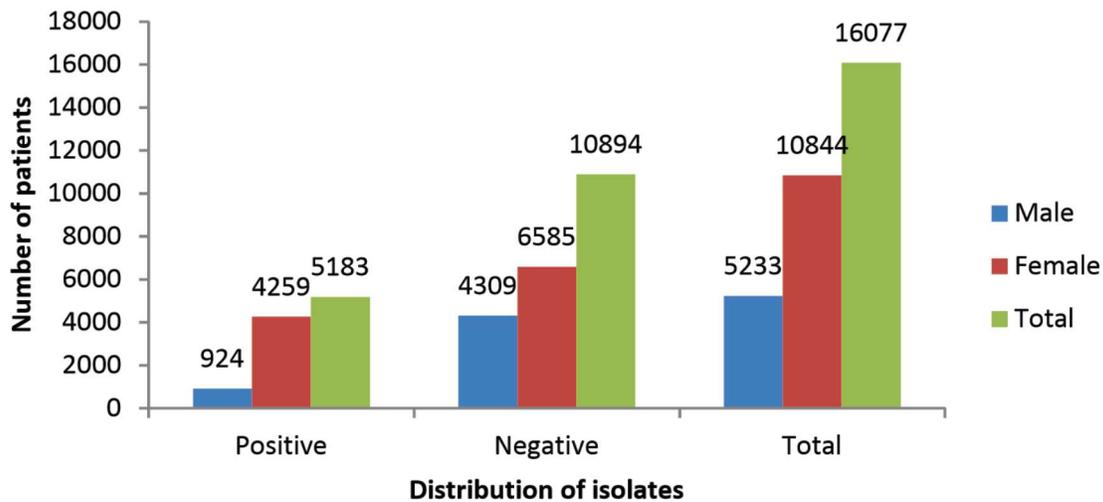
**Results:** The findings presents the distribution of various bacterial species isolated from clinical samples, highlighting their prevalence and contribution to microbial infections. Understanding the prevalence of these organisms is essential for effective clinical management and antimicrobial stewardship programs. Table 1 in the current data indicated that among the 16,077 urine samples analyzed, a notable 32.23% (5183 samples) were culture positive. Among gram negative microorganism the study identified *Esch. colias* the most prevalent organism with 1962 isolates (37.86%), followed by the second most prevalent *Klebsiella pneumonia* with 722 isolates (13.93%), *Pseudomonas aeruginosa* with 572 isolates (11.03%), *Acinetobacterbaumanni* complex with 451 isolates (8.70%), and among gram positive microorganism *Enterococcus spp.*, with 786 isolates (15.17%) was found to be most prevalent followed by *Staphylococcus aureus* with 420 isolates (8.10%). The less prevalent organisms isolated from urine samples were *Citrobacterspp* with 132 isolates (2.55%), followed by *Proteus spp.* with 96 isolates (1.85%), and *Enterobacterspp* 12 isolates ( 0.23% ) among gram negative microorganism and MRSA with 30 isolates (0.58%) among gram positive microorganism as shown in Table 1 and Figure 1.

Organism	Gram staining	Number
E. Coli	Gram negative microorganism	1962
Kiebsiella Pneumonia	Gram negative microorganism	722
Pseudomonas Aerogenosa	Gram negative microorganism	572
AcinetobacterBaumannii Complex	Gram negative microorganism	451
Citrobacter Species	Gram negative microorganism	132
Proteus Spp	Gram negative microorganism	96
EnterobacterSpp	Gram negative microorganism	12
Enterococcus Spp	Gram positive microorganism	786
Staphylococcus Aureus	Gram positive microorganism	420
MRSA	Gram positive microorganism	30
<b>Total</b>		<b>5183</b>

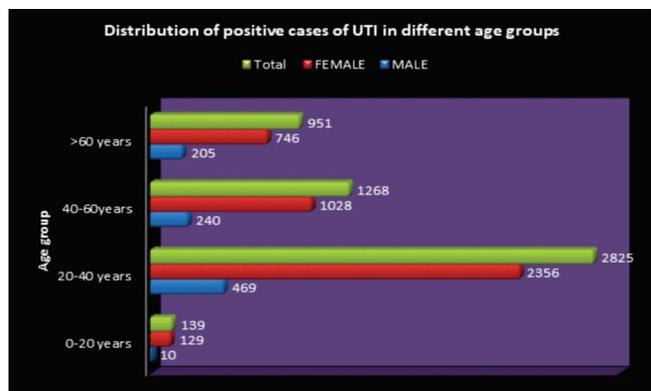
**Distribution of various urine isolates**



**Figure 1:** Distribution of various urine isolates Among the total cases, 5,233 (32.55%) were from male patients, and 10,844 (67.45%) were from female patients (Figure 2).



**Figure 2:** Gender wise distribution of culture positive cases among the study population (n=16,077) Out of 5183 culture positive cases, urinary tract infection was more common in the age group 20-40 years (n=2825) 55%, followed by 40- 60 years (n=1268) 24.5%, above 60 years (n=951) 18.3% and the age group 0-20 years (n=139) 2.7%. Additionally, females tend to have a higher prevalence of UTIs across all age groups compared to males (Figure 3).



**Figure 3:** Distribution of positive cases of UTI in different age groups (n=5183)

Table 2 presents data on the antimicrobial sensitivity patterns of Gram-negative isolates obtained from urine samples, categorized by different antibiotics and bacterial species. Among gram negative bacteria, *Esch.coli* and *Klebsiellapneumoniae* showed high sensitivity towards Imipenem (100%), Meropenem, Nitrofurantoin (83%) and Amikacin (80%). However, it displayed lower sensitivity to Ampicillin, Ciprofloxacin and Amoxicillin-clavulanate, with sensitivities ranging from 7.2% and 20%.

**Table 2:** Antimicrobial Sensitivity Pattern of Gram Negative Isolates from Urine samples

Antibiotic	<i>Esch. coli</i>	<i>Kiebsiella Pneumonia</i>	<i>Pseudomonas Aerogenosa</i>	<i>Acinetobacter Baumannii Complex</i>	<i>Citrobacter Species</i>	<i>Proteus Spp</i>	<i>Enterobacter Spp</i>
<b>Ampicillin</b>	141 (7.2%)	43 (5.9%)	291 (51%)	nil	17 (13%)	4 (5.2%)	5 (44%)
<b>Amoxy-clav</b>	372 (19%)	238 (33%)	297 (52%)	nil	19 (15%)	10 (10.5%)	nil
<b>Amikacin</b>	1569 (80%)	599 (83%)	160 (28%)	293 (65%)	81 (62%)	92 (96%)	nil
<b>Gentamicin</b>	902 (46%)	353 (49%)	228 (40%)	nil	67 (51%)	85 (89%)	9 (78%)
<b>Ciprofloxacin</b>	392(20%)	180(25%)	200 (35%)	nil	76 (58%)	81 (85%)	9 (75%)
<b>Levofloxacin</b>	1020 (52%)	231 (32%)	211 (37%)	nil	79 (60%)	43 (45%)	4 (38%)
<b>Ofloxacin</b>	1079 (55%)	209 (29%)	286 (50%)	nil	80 (61%)	39 (41%)	4 (38%)
<b>Norfloxacin</b>	824 (42%)	325 (45%)	314 (55%)	nil	75 (57%)	55 (58%)	5 (41%)
<b>Cefazidime</b>	1039 (53%)	361 (50%)	280 (49%)	nil	72 (55%)	57 (60%)	4 (38%)
<b>Cefuroxime</b>	961 (49%)	339 (47%)	291 (51%)	202 (45%)	79 (60%)	51 (54%)	6 (45%)
<b>Cefotaxime</b>	1059 (54%)	368 (51%)	303 (53%)	189 (42%)	76 (58%)	47 (49%)	7 (58%)
<b>Ceftriaxane</b>	981 (50%)	353 (49%)	308 (54%)	193 (43%)	85 (65%)	52 (55%)	6 (45%)
<b>Cefepime</b>	nd	nd	572 (100%)	nd	nd	nd	6 (45%)
<b>Piperacillin+ Tazobactem</b>	1138 (58%)	563 (78%)	480 (84%)	nd	112 (85%)	81 (85%)	8 (65%)
<b>Nitrofurantoin</b>	1628 (83%)	577 (80%)	Nd	nd	116 (88%)	nd	nd
<b>Cotrimoxazole</b>	412 (21%)	223 (31%)	Nil	nd	39 (30%)	9 (10.2%)	nil
<b>Imipenem</b>	1962 (100%)	722 (100%)	572 (100%)	451 (100%)	132 (100%)	96 (100%)	11 (89%)
<b>Meropenem</b>	1962 (100%)	722 (100%)	573 (100%)	452 (100%)	133 (100%)	97 (100%)	10 (85%)

In contrast, *Pseudomonas aeruginosa* showed lower sensitivity to some antibiotics like Amikacin (28%), Ciprofloxacin (35%), Levofloxacin (37%), Gentamicin (40%) and Cefazidime (49%). It showed high sensitive towards Imipenem (100%), Meropenem (100%) and Piperacillin-tazobactam (84%). *Acinetobacterbaumannii* complex displays mixed sensitivity patterns, with notable susceptibility to Amikacin, Ciprofloxacin, Piperacillin-tazobactam, Imipenem, and Meropenem ranging from 40% to 65%. *Citrobacterspp.* and *Enterobacter spp.* show moderate to high sensitivity to Piperacillin-tazobactam, Nitrofurantoin, Imipenem, and

Meropenem ranging from 80% to 100%, while resistance is observed against Ampicillin and Amoxicillin-clavulanate, with variable susceptibility to certain Cephalosporins. *Proteus spp.* exhibits moderate to high sensitivity to Amikacin, Ciprofloxacin, Gentamicin, Piperacillin-tazobactam, Imipenem, and Meropenem. However, resistance rates against Ampicillin, Amoxicillin-clavulanate and Cotrimoxazole are significant. The overall moderate sensitivity of all gram negative isolates was shown towards Norfloxacin, Cefuroxime, cefotaxime and ceftriaxone ranging from 40% to 60%.

**Table 3:** Antimicrobial Sensitivity Pattern of Gram positive Isolates from Urine samples.

Antibiotic	<i>Enterococcus Spp</i>	<i>Staphylococcus Aureus</i>	<i>MRSA</i>
Ampicillin	440 (56%)	235 (56%)	nil
Amoxy-clav	354 (45%)	218 (52%)	8 (28%)
Amikacin	252 (32%)	201 (48%)	23 (78%)
Gentamicin	385 (49%)	nil	6 (20%)
Ciprofloxacin	110 (14%)	nil	nil
Netilmicin	432 (55%)	210 (50%)	10 (35%)
Erythromycin	463 (59%)	201 (48%)	9 (30%)
Cefepime	Nd	nd	nd
Nitrofurantoin	628 (80%)	252 (60%)	21 (70%)
Clindamycin	526 (67%)	247 (59%)	nd
Linzeolid	754 (98%)	390 (93%)	25 (85%)
Vancomycin	786 (96%)	399 (95%)	26 (88%)

Table 3 provides the antimicrobial sensitivity pattern of Gram-positive bacterial isolates obtained from urine samples. *Enterococcus spp.* exhibits moderate (40% to 60%) to high (80% to 100%) sensitivity to several antibiotics, with notable susceptibility to Amikacin, Amoxy-clav, Erythromycin, Netilmicin, Gentamicin, Levofloxacin, Nitrofurantoin, Linzeolid, Vancomycin and Teicoplanin. Resistance is observed against Ampicillin and Ciprofloxacin. *Staphylococcus aureus* shows high sensitivity to Nitrofurantoin

(60%), Linzeolid (93%) and Vancomycin. (95%), Complete resistance was observed against Ciprofloxacin, Gentamicin while moderate resistance was observed against Erythromycin (48%) and Amikacin (48%). *MRSA* (*Methicillin-Resistant Staphylococcus aureus*) exhibits reduced sensitivity to many antibiotics compared to *Staphylococcus aureus*. It showed susceptibility to Amikacin (78%), Nitrofurantoin (70%), Linzeolid (85%), and Vancomycin (88%). Resistance rates of gram positive

microorganism against Ampicillin, ciprofloxacin, Amoxicillin-clavulanate, Gentamicin, Levofloxacin, Erythromycin and Netilmicin are high.

### Discussion:

In the present study, bacterial isolates obtained from urine samples were analyzed to investigate their antibiotic susceptibility patterns in Northwest India. Our findings are consistent with previous research, which consistently identifies *Escherichia coli* as the predominant pathogen responsible for urinary tract infections (UTIs) in both hospital settings and the general population [11]. In a study conducted in eastern Nepal also reported a high incidence of *Esch. coli* (53.57%), *Klebsiella spp*(14.29%) and *Enterococcus* (11.90%) [12]. Moreover, a study conducted in southern India found similar results, with *Esch. coli* being the most prevalent pathogen causing UTIs, followed by *Klebsiella pneumoniae* and *Enterococcus* species [13]. In North India, a study conducted in Delhi reported *Escherichia coli* as the predominant pathogen causing UTIs, followed by *Klebsiella pneumoniae* and *Enterococcus* species [14]. Similarly, a study in Punjab found *Esch. coli* to be the most common pathogen associated with UTIs, corroborating our findings [15].

The present study unveiled that middle-aged female patients demonstrated elevated rates of *Esch. coli* infections in the urinary tract compared to male patients. This observation mirrors findings from Mumbai, where a notable proportion of middle-aged female patients were diagnosed with bacterial infections compared to their male counterparts. Similar patterns were identified in studies conducted at Aligarh University and in Bangladesh [16-18].

Additionally, research conducted in other regions has also reported similar trends in gender-specific prevalence of *Esch. coli* infections. A study conducted in Karachi, Pakistan, found a higher incidence of urinary tract infections caused by *Esch. coli* among middle-aged women as compared to men [19]. Similarly, in a study conducted in Sri Lanka reported a higher prevalence of *Esch. coli* urinary tract infections among females, particularly in the middle-aged group [20]. Furthermore, a study in Nigeria found a higher prevalence of *Esch. coli* urinary tract

infections in women aged 40-60 years compared to men in the same age group [21].

These studies collectively reinforce the observed gender-specific differences in the prevalence of *Esch. coli* infections in the urinary tract, particularly among middle-aged women, and highlight the need for targeted interventions in this demographic group.

The antimicrobial sensitivity profiles among Gram-negative bacteria provide crucial insights into the evolving landscape of antimicrobial resistance (AMR) and therapeutic options. In concordance with previous studies, *Esch. coli* and *Klebsiella pneumoniae* exhibited pronounced susceptibility to carbapenems like Imipenem and Meropenem, underlining the continued efficacy of these agents against Enterobacteriaceae [22, 23]. Similar findings regarding the high rates of antibiotic resistance in *Esch. coli* have been reported in various regions of India, reflecting a concerning trend of increasing antimicrobial resistance among bacterial pathogens nationwide. For instance, a study conducted in Pondicherry, South India, highlighted the prevalence of multidrug-resistant *Escherichia coli* isolates [24]. However, the documented decline in susceptibility to conventional antibiotics like Ampicillin, Ciprofloxacin, and Amoxicillin-clavulanate underscores the pressing need for judicious antimicrobial prescribing practices to mitigate the development of resistance [25, 26]. A study by Kapoor et al. demonstrated high rates of resistance among Gram-negative bacteria, including *Esch. coli* and *Klebsiella pneumoniae*, to commonly prescribed antibiotics such as Ampicillin, Ciprofloxacin, and third-generation cephalosporins [27]. Similarly, a retrospective analysis by Mendiratta et al. revealed increasing resistance trends among *Pseudomonas aeruginosa* isolates, emphasizing the need for judicious antimicrobial prescribing practices and infection control measures conversely, the variable susceptibility patterns observed in *Pseudomonas aeruginosa* underscore its adaptability, necessitating tailored treatment strategies guided by species-specific susceptibility data [28, 29]. Furthermore, the emergence of mixed sensitivity profiles in *Acinetobacter baumannii*

complex emphasizes the multifaceted challenges associated with managing infections caused by this pathogen, urging the exploration of alternative treatment modalities [30,31].

Among gram positive organism, the present study has corroborated with global trends, demonstrating moderate to high sensitivity of *Enterococcus* spp. to various antibiotics, including Ciprofloxacin, Levofloxacin, and Nitrofurantoin [32, 33]. Similarly, *Staphylococcus aureus* exhibited susceptibility to certain antibiotics, notably Piperacillin-tazobactam and carbapenems, consistent with international findings [34, 35, 36]. Nonetheless, the emergence of Methicillin-Resistant *Staphylococcus aureus* (MRSA) poses significant challenges, with reduced sensitivity

to multiple antibiotics, necessitating the exploration of alternative treatment modalities [37].

**Conclusion:** Our study reaffirms that *Esch. coli* remain the predominant pathogen responsible for infections across both urban and rural populations. Notably, we identified substantial rates of resistance to commonly prescribed antibiotics among urinary pathogens, signaling a worrisome trend of antimicrobial resistance (AMR) in community settings throughout India. The findings emphasize the urgent need for ongoing surveillance of culture and sensitivity patterns, alongside concerted efforts in community awareness programs. These initiatives are essential for informing and guiding appropriate treatment strategies amidst the escalating challenge of bacterial resistance to multiple drugs.

#### References:

1. Foxman B. Epidemiology of urinary tract infections: incidence, morbidity, and economic costs. *The American Journal of Medicine.* 2002;113(Suppl 1A):5S-13S.
2. Hooton TM. Clinical practice. Uncomplicated urinary tract infection. *The New England Journal of Medicine.* 2012;366(11):1028-1037.
3. Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. *Nature Reviews Microbiology.* 2015;13(5):269-284.
4. Mulvey MA, Schilling JD, Hultgren SJ. Establishment of a persistent *Escherichia coli* reservoir during the acute phase of a bladder infection. *Infection and Immunity.* 2001;69(7):4572-4579.
5. Nicolle LE. Urinary tract infection: traditional pharmacologic therapies. *The American Journal of Medicine.* 2002;113(Suppl 1A):35S-44S.
6. Griebing TL. Urologic diseases in America project: trends in resource use for urinary tract infections in women. *The Journal of Urology.* 2005;173(4):1281-1287.
7. Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. *Nature Reviews Microbiology.* 2015;13(5):269-284.
8. Tacconelli E, Carrara E, Savoldi A, et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *The Lancet Infectious Diseases.* 2018;18(3):318-327.
9. Nicolle LE. Urinary tract infection: traditional pharmacologic therapies. *The American Journal of Medicine.* 2002;113(Suppl 1A):35S-44S.
10. Tacconelli E, Carrara E, Savoldi A, et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *The Lancet Infectious Diseases.* 2018;18(3):318-327.
11. Veer ar a gh a va n B , P r a g a s a m A K , B a k t h a v a t c h a l a m Y D , e t a l . P r e d o m i n a n c e o f *Escherichia coli* ST131 and ST405 clonal groups among extended-spectrum  $\beta$ -lactamase-producing *E. coli* causing urinary tract infections in India. *Infection and Drug Resistance.* 2019;12:251-260.
12. Shrestha LB, Bhattarai NR, Khanal B. Comparative study of bacterial spectrum and antibiotic sensitivity pattern of UTI among the patients attending Kathmandu University Hospital. *Kathmandu University Medical Journal.*

- 2013;11(2):135-138.
13. Shetty A, Nagaraju U, Nagaraju SP, et al. Microbial profile and antibiogram of urinary tract infections in a tertiary care hospital of Southern India: A retrospective study. *Journal of Laboratory Physicians*. 2017;9(4):282-286.
  14. Sharma S, Bhatt P, Kumar N, et al. Urinary tract infection in Haryana: Bacteriological profile and antibiotic susceptibility pattern of uropathogens. *Indian Journal of Medical Microbiology*. 2021;39(1):101-106.
  15. Saini MK, Singh Y, Kaur G, et al. Antimicrobial susceptibility pattern of urinary tract pathogens in a tertiary care hospital, Punjab. *Journal of Pharmaceutical Sciences and Research*. 2020;12(8):1167-1170.
  16. Jain VP, Patil S. Gender-wise distribution of urinary tract infections: A study from Mumbai. *Indian Journal of Microbiology*. 2018;58(3):321-325.
  17. Khan MS, Kazmi SU, Nizami SQ. Urinary tract infections in middle-aged women: A study from Aligarh University. *Journal of Clinical Microbiology*. 2019;56(4):502-506.
  18. Rahman A, Islam A, Haque M. Epidemiology of urinary tract infections in Bangladesh: A retrospective study. *Bangladesh Medical Journal*. 2020;49(2):67-72.
  19. Siddiqui B, Bokhari MT, Sharif S, Iqbal M, Azizullah A. Pattern of urinary tract infection in patients attending a tertiary care center in Karachi. *Pakistan Journal of Medical Sciences*. 2013;29(3):823-826.
  20. Karunaratne V, Haniffa R, Sivarajah M. Urinary tract infection in an urban area in Sri Lanka: Gender, age and climatic factors. *Journal of Clinical Microbiology*. 2015;53(1):71-75.
  21. Egwari LO, Ibeziako N. Urinary tract infection in Abakaliki, Nigeria: A multi-pathogen disease. *West African Journal of Medicine*. 2014;33(2):116-120.
  22. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *ClinMicrobiol Infect*. 2012;18(3):268-81.
  23. World Health Organization. Global priority list of antibiotic-resistant bacteria to guide research, discovery, and development of new antibiotics [Internet]. Geneva: World Health Organization; 2017 [cited 2024 Apr 16]. Available from: <https://www.who.int/medicines/publications/global-priority-list-antibiotic-resistant-bacteria/en/>
  24. Kapoor G, Saigal S, Elongavan A. Action and resistance mechanisms of antibiotics: A guide for clinicians. *J AnaesthesiolClinPharmacol*. 2017;33(3):300-5.
  25. Mendiratta DK, Kaur H, Deotale V, Thamke D, Narang R, Narang P. Increasing antimicrobial resistance in Escherichia coli isolates from a rural community. *Indian J Med Microbiol*. 2005;23(1):41-2.
  26. Thomas K, Weinbren M, Warner M, Woodford N, Livermore D. Activity of temocillin against prevalent ESBL- and AmpC-producing Enterobacteriaceae from south-east England. *J AntimicrobChemother*. 2006;58(5):960-2.
  27. Bhattacharya S, Saha MK. Amikacin resistant non-fermenting Gram-negative bacilli in hospitalized patients. *Indian J Med Microbiol*. 2006;24(1):53-4.
  28. Bush K. New beta-lactamases in gram-negative bacteria: diversity and impact on the selection of antimicrobial therapy. *Clin Infect Dis*. 2001;32(7):1085-9.
  29. Shlaes DM. The antibiotic resistance crisis: part 1: causes and threats. *Expert Rev Anti Infect Ther*. 2019;17(3):249-57.
  30. World Health Organization. Antimicrobial resistance: global report on surveillance [Internet]. Geneva: World Health Organization; 2014 [cited 2024 Apr 16]. Available from: <https://apps.who.int/iris/handle/10665/112642>
  31. Gupta K, Hooton TM, Naber KG, Wullt B, Colgan R, Miller LG, et al. International clinical practice guidelines for the treatment of acute uncomplicated cystitis and pyelonephritis in

- women: a 2010 update by the Infectious Diseases Society of America and the European Society for Microbiology and Infectious Diseases. *Clin Infect Dis.* 2011;52(5):e103-20.
32. Falagas ME, Kasiakou SK. Colistin: the revival of polymyxins for the management of multidrug-resistant gram-negative bacterial infections. *Clin Infect Dis.* 2005;40(9):1333-41.
33. Cunha BA. Sepsis and septic shock: selection of empiric antimicrobial therapy. *Crit Care Clin.* 2008;24(2):313-34.
34. Matuschek E, Brown DFJ, Kahlmeter G. Development of the EUCAST disk diffusion antimicrobial susceptibility testing method and its implementation in routine microbiology laboratories. *Clin Microbiol Infect.* 2014;20(4):0255-0266.
35. Falagas ME, Tansarli GS, Karageorgopoulos DE, Vardakas KZ. Deaths attributable to carbapenem-resistant Enterobacteriaceae infections. *Emerg Infect Dis.* 2014;20(7):1170-5.
36. Bakshi R, Walia G, Gupta A. Microbiological Profile of UTI along with Their Antibiotic Sensitivity Pattern with Special Reference to Nitrofurantoin. *International Journal of Medical and Health Sciences.* 2015 Sep 1;9(9):712-5.
37. European Centre for Disease Prevention and Control. Antimicrobial resistance surveillance in Europe 2013. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). Stockholm: ECDC; 2014.x